

**Recent Progress in Medicinal
Plants**

Volume 21

***Phytopharmacology
and
Therapeutic Values III***

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ISBN : 1-9336991-1-6
SERIES ISBN : 0-9656038-5-7

2008



Stadium Press LLC, U.S.A.

P.O. Box-722200, Houston, Texas-77072, USA
Tel.: 713-541-9400; Fax : 713-541-9401
E-mail : studiumpress@studiumpress.com

Genus *Baccharis* (Asteraceae): A Review of Chemical and Pharmacological Studies

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Abstract

The genus Baccharis belongs to the family Asteraceae, tribe Astereae and subtribe Baccharidinae, and it is distributed from the United States to Argentina. It contains approximately 400 species, many of them economically important as medicines, ornamentals and sources of essential oil. Some species, popularly called carqueja, show caulinar expansions or wings which constitute cladodes. In folk medicine, these species are considered stomachic and diuretic. Chemical investigations have focused on compounds of the essential oils, diterpenoids, flavonoids and macrocyclic trichothecenes. Pharmacological essays have demonstrated their antiinflammatory, antimicrobial, antioxidant and cytotoxic activities.

Key words : Asteraceae, *Baccharis*, Chemistry, Essential oil, Pharmacology, Trichothecene

Introduction

The Asteraceae Dumort., also known as Compositae Giseke, are one of the major families of Asterales and comprehend approximately 1,500 genera and 23,000 species, found especially in temperate or tropical montane regions and dry habitats (Judd *et al.*, 1999). The family includes herbs, shrubs, lianas and rarely trees. About 98 % of the genera are herbaceous (Barroso, 1991; Joly, 1998). The evolutionary success of the taxon may be attributed to the array of secondary metabolites, such as terpenoid essential oils, caffeic

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acid derivatives, flavonoids, alkaloids and the defense combination of polyacetylenes and sesquiterpene lactones (Emerenciano *et al.*, 1986; Mesquita *et al.*, 1986; Cronquist, 1988). Flavonoids, particularly flavones, are taxonomically relevant at the tribal and subtribal level (Emerenciano *et al.*, 2001). Among the major genera, *Baccharis* contains about 400 species (Judd *et al.*, 1999).

Systematic aspects

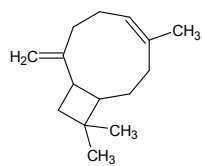
The genus *Baccharis* belongs to the tribe Astereae and subtribe Baccharidinae, and it is distributed from the United States to Argentine, mainly in South America. Many species have been found in Brazil and Andes, and therefore this region is viewed as the probable centre of distribution of the genus. In the Southwest of Brazil, 120 members have been reported. For systematic diagnosis, the leaf morphology and inflorescence type are chiefly important (Barroso & Bueno, 2002).

Morphological characters

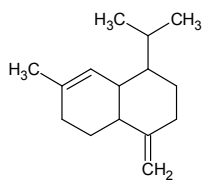
Baccharis members are perennial shrubs, dioecious, that attain height of 50 cm to 4 m and in general, have alternate leaves and cymose inflorescences. Female plants bear female flowers and, in male plants, the flowers are morphologically hermaphrodite and functionally male owing to the sterile gynoeceum (Ariza-Espinar, 1973; Barroso & Bueno, 2002). Many species show caulinar expansions or wings which constitute cladodes, considered modified stems that play a photosynthetic role. These species are popularly called carqueja in Brazil and Argentine, and exhibit two or three winged cladodes. Some works have dealt with anatomical characters of aerial vegetative organs for pharmacognostic purposes and have pointed out features that many species share, such as anomocytic stomata, glandular and non-glandular trichomes, calcium oxalate crystals, secretory ducts and an evident endodermis or starch sheath (Ariza-Espinar, 1973; Barroso, 1976; Chicourel *et al.*, 1997; Oliveira & Bastos 1998; Cortadi *et al.*, 1999; Gianello *et al.*, 2000; Ortins & Akisue 2000; Budel *et al.*, 2003a; 2003b; 2004a; 2004b). A review on the subject has been recently published (Budel *et al.*, 2005).

Ethnobotanical uses

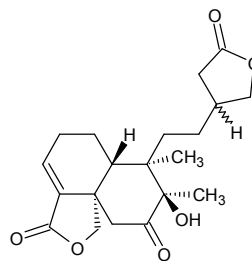
Baccharis is economically important for providing medicinal plants, ornamentals, hosts of pollinating insects and galls, sources of essential oil and for protecting areas from soil erosion (Barroso, 1976; Matzenbacher, 1985; Carneiro & Fernandes, 1996; Alonso, 1998; Oliveira & Bastos, 1998; Castro & Ferreira, 2001; Midorikawa *et al.*, 2001; Kumazawa *et al.*, 2003; Santos *et al.*, 2003). The winged species, known as carqueja, are used as diuretic and stomachic in folk medicine. Among them, *B. articulata* (Lam.) Pers. (Fig 1), *B. crispa* Spreng., *B. gaudichaudiana* DC. (Fig 2), *B.*



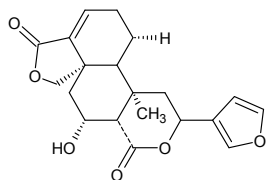
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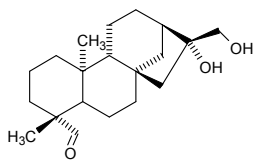
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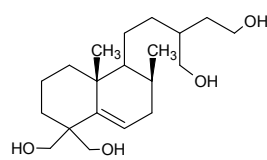
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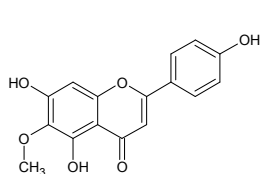
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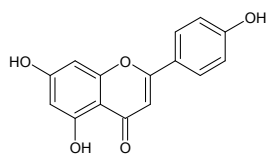
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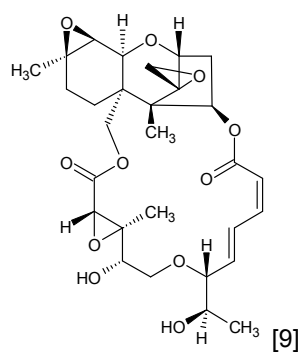
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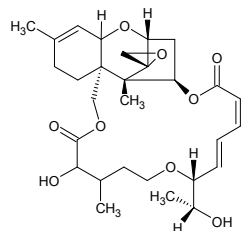
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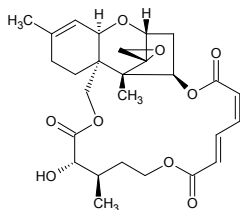
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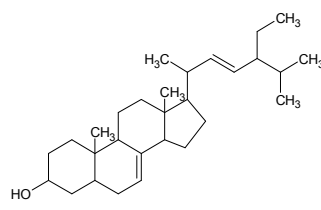
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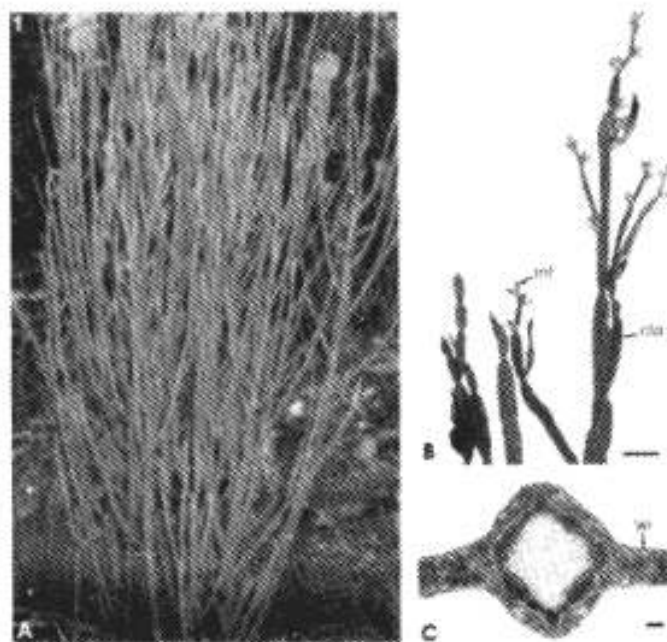


Fig 1. *Baccharis articulata* (Lam.) Pers.: **A.** Aspect of cladodes; **B.** Detail of inflorescences (inf) and cladodes (cla); **C.** Cross-section of a cladode with two wings (wi).

microcephala Baker, *B. myriocephala* DC., *B. pseudovillosa* I.L. Teodoro and J.E. Vidal, *B. sagittalis* (Less.) DC., *B. stenocephala* Baker, *B. trimera* (Less.) DC., *B. usterii* Heering and *B. vincaefolia* Baker are found (Ariza-Espinar, 1973; Corrêa, 1984; Barroso & Bueno, 2002; Budel *et al.*, 2005). Apart from carquejas, *B. dracunculifolia* DC. (Fig 3) is also a medicinal plant whose leaves are used to treat gastric disorders (Mors *et al.*, 2000) and yield a prized essential oil in perfumery (Craveiro *et al.*, 1981).

Chemical and pharmacological studies

This review collates data about investigations which focus on chemical and pharmacological essays of *Baccharis*. A summary of the different studies is presented in Tables 1 and 2.

This genus is a source of essential oil and various works have dealt with the identification of compounds, such as α -cadinene, camphene, carquejil acetate, carquejol, β -caryophyllene[1], limonene, γ -muurolene [2], α -pinene, β -pinene and spathulenol (Siqueira *et al.*, 1985; Weyerstahl *et al.*, 1990; 1996; Ferracini *et al.*, 1995; Loayza *et al.*, 1995; Zunino *et al.*, 1997; 1998; 2000; Frizzo *et al.*, 2001; Agostini *et al.*, 2005). Diterpenoids and flavonoids are the major chemical groups found in *Baccharis*. The former

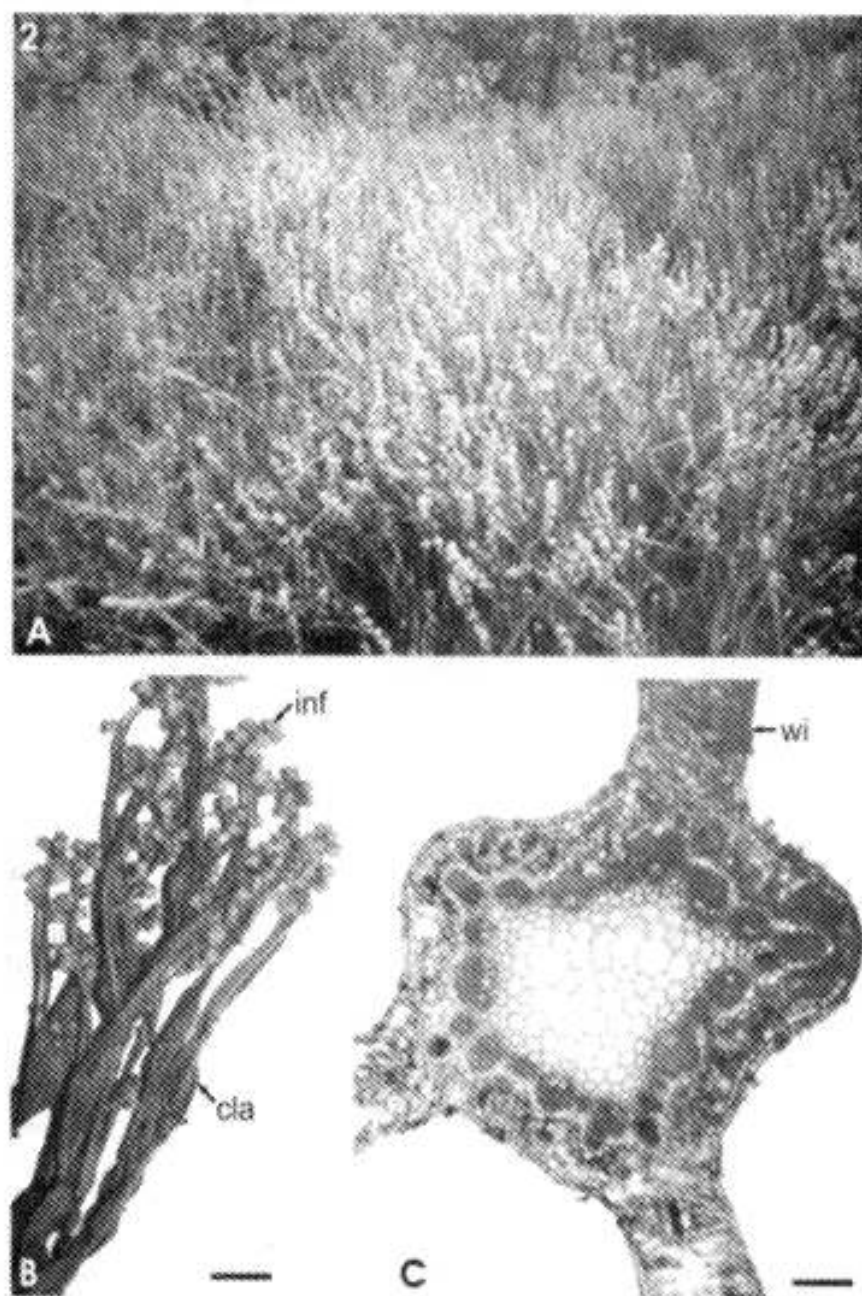


Fig 2. *Baccharis gaudichaudiana* DC.: **A.** Aerial vegetative and reproductive organs; **B.** Inflorescences (inf) & cladodes (cla); **C.** Cross-section of a cladode with three wings (wi).



Fig 3. *Baccharis dracunculifolia* DC.: **A.** Aspect of the plant; **B.** Apical stems bearing inflorescences (inf) & leaves (le).

includes clerodanes, *e.g.* 8 β -hydroxy-7-oxo-ent-cleroda-3-en-15,18-diacid-16,19-dilactone [3] and 7 α -hydroxybacchotricuneatin A [4], kauranes, *e.g.* 14 α -17-dihydroxy-ent-kauran-19-al [5], and labdanes, *e.g.* gaudichaudol A [6] (Zdero *et al.*, 1991; Dai *et al.*, 1993; Wachter *et al.*, 1999; Cifuentes *et al.*, 2001a; Hikawczuk *et al.*, 2002; Akaike *et al.*, 2003). Of the different types of flavonoids found in the genus, the flavones are the most diverse group (Bandoni *et al.*, 1978; Sharp *et al.*, 2001; Moreira *et al.*, 2003a). Some of the biologically active flavones isolated are hispidulin [7] and apigenin [8] (Soicke & Leng-Peschlow 1987; Moreira *et al.*, 2003b).

Moreover, macrocyclic trichothecenes have been recorded in *Baccharis*, *e.g.* *B. coridifolia* DC., *B. megapotamica* Spreng. and *B. artemisioides* Hook. and Arn. in Hook. (Jarvis *et al.*, 1988; 1991; Bergmann *et al.*, 1992; Rizzo *et al.*, 1997; Varaschin & Alessi, 2003). This group of chemicals has antiviral properties and has been tested for cancer treatment. Baccharin [9], a trichothecene obtained from *B. megapotamica*, and roridins, *e.g.* roridin A [10], and verrucarins, *e.g.* verrucarins A [11], isolated from *B. coridifolia* are considered bioactive molecules against various tumour cell lines.

Table 1. Chemical studies of *Baccharis* species.

<i>Baccharis</i> species	Results	References
<i>B. sarothroides</i> A. Gray	Isolation of two flavonols: 3',4'-dimethoxy-3,5,7-trihydroxyflavone and centaureidine	Kupchan & Baugers Schmidt (1971)
<i>B. crispa</i> Spreng.	Identification of 5-hydroxy-7,4'-dimethoxyflavone and 5,3'-dihydroxy-7,4'-dimethoxyflavone	Bandoni <i>et al.</i> (1978)
<i>B. articulata</i> (Lam.) Pers., <i>B. crispa</i> Spreng., <i>B. gilliesii</i> A. Gray, <i>B. myrtilloides</i> Griseb., <i>B. pingraea</i> DC., <i>B. ulicina</i> Hook. and Arn.	Characterization of jaceosidin and oleamolic acid in <i>B. gilliesii</i> , jaceosidin and lupool in <i>B. myrtilloides</i> , chondrillasterol [12] in <i>B. ulicina</i> and <i>B. pingraea</i> , acetoin and cirsimarin in <i>B. articulata</i> , salvigenin and 7,4'-dimethylapigenin in <i>B. articulata</i> and <i>B. crispa</i>	Gianello & Giordano (1984)
<i>B. chilco</i> Kunth, <i>B. eggersii</i> Hieron., <i>B. hutchinsonii</i> Cuatrec., <i>B. microcephala</i> Baker, <i>B. nitida</i> (Ruiz and Pav.) Pers., <i>B. phyllonoides</i> Kunth, <i>B. scoparia</i> (L.) Sw.	Isolation of five clerodanes, three labdane derivatives and a <i>nor</i> -diterpene, α hydroperoxide derived from 4 α -hydroxygermacra-1(10),5-diene	Bohlmann <i>et al.</i> (1985)
<i>B. articulata</i> (Lam.) Pers., <i>B. trimera</i> (Less.) DC.	Principal constituents identified, respectively: carquejil acetate (45.0% and 67.0%), camphene (23.0% and 3.7%), carquejol (9.0% and 18.5%), β -pinene (5.0% and 3.0%) and α -pinene (3.0% and 0.5%)	Siqueira <i>et al.</i> (1985)
<i>B. incurum</i> (Wedd.) Cuatrec.	Isolation of five neo-clerodane diterpenes: a kolavane derivative, two clerodane lactones, 7 α -hydroxybacchotricuneatin A [4] and 1 α -acetylbacchotricuneatin A	Givovich <i>et al.</i> (1986)
<i>B. trimera</i> (Less.) DC.	Identification of quercetin, luteolin, nepetin, hispidulin [7] and apigenin [8]	Soicke & Leng-Peschlow (1987)
<i>B. cordifolia</i> DC.	Isolation of macrocyclic trichothecenes similar to <i>Myrothecium</i> fungus	Jarvis <i>et al.</i> (1988)

Baccharis species	Results	References
<i>B. bigelovii</i> A. Gray, <i>B. halimifolia</i> L., <i>B. heterophylla</i> Kunth, <i>B. neglecta</i> Britton, <i>B. potosina</i> A. Gray, <i>B. pteronioides</i> DC., <i>B. salicifolia</i> (Ruiz and Pav.) Pers., <i>B. salicina</i> Torr. and A. Gray, <i>B. sarothroides</i> A. Gray, <i>B. thesioides</i> Kunth	Isolation of seven <i>ent</i> -clerodanes and seven labdanes, three of them glycosides, a kaurane and a lactone derived from geranylinalol	Jakupovic <i>et al.</i> (1990)
<i>B. paniculata</i> DC. <i>B. dracunculifolia</i> DC. <i>B. gaudichaudiana</i> DC.	Isolation of two <i>ent</i> -labdanes Identification of iso-humbertiol and dracunculifolol Characterization of gaudichaudiosides A-E	Faini & Castillo (1990) Weyerstahl <i>et al.</i> (1990) Fullas <i>et al.</i> (1991)
<i>B. cordifolia</i> DC., <i>B. megapotamica</i> Spreng. <i>B. petiolata</i> DC., <i>B. santelicensis</i> Phil.	Isolation of macrocyclic trichothecenes Isolation of dimeric sesquiterpenes and five labdane derivatives in <i>B. petiolata</i> and furoclerodanes and phenolics in <i>B. santelicensis</i>	Jarvis <i>et al.</i> (1991) Zdero <i>et al.</i> (1991)
<i>B. gaudichaudiana</i> DC.	Identification of gaudichaudioside F	Fullas <i>et al.</i> (1992)
<i>B. articulata</i> (Lam.) Pers., <i>B. cylindrica</i> (Lam.) Pers., <i>B. microcephala</i> Baker, <i>B. trimera</i> (Less.) DC., <i>B. ustarii</i> Heering	Characterization of triterpenes in all species, as well as flavonoids, polyphenols and cardiac glycosides in <i>B. articulata</i> , saponins in <i>B. cylindrica</i> , alkaloids, flavonoids and saponins in <i>B. microcephala</i> , polyphenols, flavonoids and saponins in <i>B. trimera</i> , flavonoids and saponins in <i>B. ustarii</i>	Bianchi <i>et al.</i> (1993)
<i>B. articulata</i> (Lam.) Pers.	Isolation of two clerodane diterpenes: 8 β -hydroxy-7-oxo- <i>ent</i> -cleroda-3-en-15,18-diacid-16,19-dilactone [3] and 15,16-epoxy-7 α ,18-dihydroxy-15-methoxy- <i>ent</i> -cleroda-3-ene	Dai <i>et al.</i> (1993)
<i>B. gaudichaudiana</i> DC.	Isolation of the labdane diterpenes gaudichaudol A	Fullas <i>et al.</i> (1994)

<i>Baccharis</i> species	Results	References
<i>B. gaudichaudiana</i> DC.	[6], B and C, the clerodane diterpenoid gaudichaudone, the clerodane artuculin acetate, hispidulin [7], apigenin [8], spathulenol and ursolic acid	Loayza <i>et al.</i> (1995)
<i>B. dracunculifolia</i> DC., <i>B. latifolia</i> (Ruiz and Pav.) Pers., <i>B. salicifolia</i> (Ruiz and Pav.) Pers.	Identification of δ -cadinene, β -caryophyllene [1], γ -elemene, germacrene-D, limonene, α -phellandrene, β -pinene, α -thujene and verbocidentatafuran among others	Weyerstahl <i>et al.</i> (1996)
<i>B. dracunculifolia</i> DC.	Constituents isolated: limonene (8.4%), β -caryophyllene [1] (6.2%), β -pinene (5.9%), δ -cadinene (5.6%), germacrene-D (3.6%), aromadendrene (3.3%), α -pinene (2.9%) and γ -muurolene [2] (2.1%)	Zumino <i>et al.</i> (1997)
<i>B. crispa</i> Spreng., <i>B. salicifolia</i> (Ruiz and Pav.) Pers.	Main constituents identified: <i>trans</i> -nerolidol (27.0%) in <i>B. crispa</i> , α -cadinol (9.4%) and germacrene-D (8.8%) in <i>B. salicifolia</i>	Wachter <i>et al.</i> (1999)
<i>B. pingraea</i> DC.	Isolation of the furolabdane angeloyl-gutierrezianolic acid, two novel diterpenoids furolabda-6,8-dien-17-oic acid and furolabda-7-en-17-oic acid and the linear diterpenoid (10E)-centipedic acid	Zunino <i>et al.</i> (2000)
<i>B. cordobensis</i> Heering	Major constituents: <i>trans</i> -nerolidol (15.8%), T-cadinol (14.7%) and cubenol (8.8%)	Nagatani <i>et al.</i> (2001 & 2002)
<i>B. dracunculifolia</i> DC.	Characterization of β -D-glucopyranose, β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranose and dracunculifosides A-J	Sharp <i>et al.</i> (2001)
<i>B. trinervis</i> Pers.	Isolation of three flavones: pectolinarigenin, salvigenin and penduletin	

Table 1. (Contd.)		
<i>Baccharis</i> species	Results	References
<i>B. uncinella</i> DC.	Quantitative determination of α -pinene (16.0%), β -pinene (15.0%), limonene (13.0%), spathulenol (10.0%), globulol (5.0%), (<i>E</i>)-nerolidol (4.0%), bicyclogermacrene (3.0%) and terpinen-4-ol (2.0.%)	Frizzo <i>et al.</i> (2001)
<i>B. medullosa</i> DC.	Isolation of two labdane diterpene glycosides	Cifuentes <i>et al.</i> (2001a)
<i>B. flabellata</i> Hook. and Arn.	Characterization of three diterpenoid derivatives: 2,19;15,16-diepoxy-neo-clerodan-3,13(16),14-trien-18- <i>oic</i> acid, 15,16-epoxy-5,10-seco-clerodan-1(10),2,4,13(16),14-pentacen-18,19- <i>olide</i> and 15,16-epoxy-neo-clerodan-1,3,13(16),14-tetraen-18,19- <i>olide</i>	Hikawczuk <i>et al.</i> (2002)
<i>B. pseudotenusifolia</i> Malag.	Isolation of oleanoic acid, α -spinasterol, hispidulin [7], naringenin, 3'-methoxy-luteolin, apigenin [8], kaempferol, eriodictiol, aromadenin, quercetin, 3'-methoxy-quercetin, quercetin-3- <i>O</i> -rhamnoside and quercetin-3- <i>O</i> -glucoside	Moreira <i>et al.</i> (2003b)
<i>B. grisebachii</i> Hieron.	Isolation of two diterpenes, eight <i>p</i> -coumaric acid derivatives and two flavones	Feresin <i>et al.</i> (2003)
<i>B. gaudichaudiana</i> DC.	Isolation of the <i>ent</i> -clerodane diterpene bacchariol	Akaike <i>et al.</i> (2003)
<i>B. ligustrina</i> DC.	Characterization of acid triterpenes and flavonoids	Moreira <i>et al.</i> (2003a)
<i>B. dracunculifolia</i> DC.	Isolation of the compounds isosakuranetin, aromadenin-4'-methyl ether, baccharis oxide, ferulic acid, dihydrocinnamic acid, 3-prenyl-4-(dihydrocinnamoyloxy)-cinnamic acid and friedelanol	Silva-Filho <i>et al.</i> (2004)
<i>B. ilirita</i> DC.	Isolation of kaurene diterpenes and flavonoids	Verdi <i>et al.</i> (2004)

Table 2. Pharmacological studies of *Baccharis* species.

<i>Baccharis</i> species	Results	References
<i>B. sarothroides</i> A. Gray	Cytotoxic activity against human carcinoma cell lines	Kupchan & Bauerschmidt (1971)
<i>B. glutinosa</i> Pers.	Antimicrobial action of pinoembrine against <i>Alternaria</i> fungus	Miyakado <i>et al.</i> (1976)
<i>B. trimera</i> (Less.) DC.	Molluscicidal effect of a diterpene lactone	Santos-Filho <i>et al.</i> (1980)
<i>B. trimera</i> (Less.) DC.	Antihepatotoxic effect	Soicke & Leng-Peschlow (1987)
<i>B. anomala</i> DC.	Mutagenic activity	Vargas <i>et al.</i> (1991)
<i>B. articulata</i> (Lam.) Pers.,	Anti-inflammatory effect	Gene <i>et al.</i> (1992)
<i>B. crispa</i> Spreng,		
<i>B. trimera</i> (Less.) DC.		
<i>B. articulata</i> (Lam.) Pers.,	Toxic effects of aqueous extract of <i>B. articulata</i>	Bianchi <i>et al.</i> (1993)
<i>B. cylindrica</i> (Lam.) Pers.,	and <i>B. usterii</i>	
<i>B. microcephala</i> Baker,		
<i>B. trimera</i> (Less.) DC.,		
<i>B. usterii</i> Heering		
<i>B. gaudichaudiana</i> DC.	Cytotoxic activities	Fullas <i>et al.</i> (1994)
<i>B. pedunculata</i> (Mill.) Cabrera	Antifungal activity against human pathogenic and phytopathogenic fungi	Rahaison <i>et al.</i> (1995)
<i>B. trimera</i> (Less.) DC.	Anti-inflammatory and analgesic activities	Gene <i>et al.</i> (1996)
<i>B. glutinosa</i> Pers.	Antimicrobial action	Verastegui <i>et al.</i> (1996)
<i>B. coridifolia</i> DC.	<i>In vitro</i> anti-oxidant and cytotoxic activities	Mongelli <i>et al.</i> (1997)
<i>B. trinervis</i> Pers.	Anti-oxidant activity	Heras <i>et al.</i> (1998)
<i>B. tucumanensis</i> Hook. & Arn.	Anti-inflammatory effect	Muschietti <i>et al.</i> (1998)
<i>B. heterophylla</i> Kunth	Antispasmodic effect	Rojas <i>et al.</i> (1999)
<i>B. teindalensis</i> Kunth,	Antiviral activity	Abad <i>et al.</i> (1999)
<i>B. trinervis</i> Pers.	Antiviral activity	Zanon <i>et al.</i> (1999)
<i>B. articulata</i> (Lam.) Pers.	Cytotoxic and DNA interaction activities	Mongelli <i>et al.</i> (2000)
<i>B. grisebachii</i> Hieron.	Bacteriostatic and bactericidal activities	Avancini <i>et al.</i> (2000)
<i>B. trimera</i> (Less.) DC.		

Table 2. (Contd.)

<i>Baccharis</i> species	Results	References
<i>B. medullosa</i> DC., <i>B. rufescens</i> Spreng.	Anti-inflammatory effect	Cifuentes <i>et al.</i> (2001b)
<i>B. notosegilla</i> Griseb.	Antimicrobial action	Cobos <i>et al.</i> (2001)
<i>B. grisebachii</i> Hieron.	Antimicrobial action	Feresin <i>et al.</i> (2001)
<i>B. coridiifolia</i> DC., <i>B. ochracea</i> Spreng.	<i>In vitro</i> cytotoxic activity against human solid tumour cell lines	Monks <i>et al.</i> (2002)
<i>B. trinervis</i> Pers.	Potent anti-HIV activity in a MTT cell proliferation assay	Sanchez-Palomino <i>et al.</i> (2002)
<i>B. conferta</i> Kunth	Antispasmodic effect	Weimann <i>et al.</i> (2002)
<i>B. articulata</i> (Lam.) Pers.	Anti-oxidant activity	Oliveira <i>et al.</i> (2003)
<i>B. heterophylla</i> Kunth	Activation of the Ca ²⁺ dependent chloride channels in <i>Xenopus laevis</i> oocytes.	Rojas <i>et al.</i> (2003)
<i>B. illinita</i> DC.	Gastroprotective effects	Baggio <i>et al.</i> (2003)
<i>B. trimera</i> (Less.) DC.	Relaxant effect on the smooth muscle of the corpus cavernosum of guinea pig	Hnatyszyn <i>et al.</i> (2003)
<i>B. grisebachii</i> Hieron.	Antimicrobial action	Feresin <i>et al.</i> (2003)
<i>B. dracunculifolia</i> DC.	Trypanocidal effect	Silva-Filho <i>et al.</i> (2004)
<i>B. trimera</i> (Less.) DC.	Anti-proteolytic and anti-hemorrhagic properties against snake venoms	Januário <i>et al.</i> (2004)
<i>B. grisebachii</i> Hieron.	Free radical scavenging action and inhibition of lipoperoxidation in erythrocytes	Tapia <i>et al.</i> (2004)
<i>B. trimera</i> (Less.) DC.	Antidiabetic activity	Oliveira <i>et al.</i> (2005)
<i>B. crispa</i> Spreng., <i>B. trimera</i> (Less.) DC., <i>B. ustera</i> Heering	Radical scavenging activity	Simões-Pires <i>et al.</i> (2005)
<i>B. latifolia</i> (Ruiz & Pav.) Pers., <i>B. obtusifolia</i> Kunth,	Anti-inflammatory effect	Abad <i>et al.</i> (2006)
<i>B. pentlandii</i> DC., <i>B. subulata</i> D. Don		

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